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# ScopeMed

# Interpreting physiology to prevent pathologies associated with obesity and diabetes

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## ABSTRACT

The approach by medicine and associated sciences dealing with the pathologies produced by obesity and Type 2 diabetes (T2D) has predominantly focused on the treatment rather than the prevention of the two health conditions. Both pharmacological and surgical approaches have been used in the treatment of both obesity and T2D. None is completely successful without incurring substantial side-effects including the option of permanent changes to the gastro-intestinal anatomy and physiology. This review puts forward for consideration two ideas: (1) That some of the failures in addressing prevention of body weight, and (2) that there is a knowledge gap in how the capacity of the pancreas and adipose tissue, key organs in the pathologies of obesity and T2D, may be modified during two important periods of tissue proliferation, the second trimester of fetal development, the pre-pubertal growth spurt, and also possibly during recovery from substantial weight loss.

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#### INTRODUCTION

Obesity and overweight conditions have risen rapidly in developed countries during the last half century. Currently, 34.9% of adult US population or about 78.6 million Americans are obese at an estimated annual medical cost of \$147 billion in 2008 US dollars [1]. Obesity affects a significant number of pathologies: hypertension [2], atherosclerosis [3], hypercoagulability of blood [4], endothelial dysfunction [5] and associated increase in the risk of coronary vascular disease and accidents [6]. Excessive fat deposition in white adipose tissue (WAT) is accompanied by progressive decline in the sensitivity of WAT to insulin action [7,8]. When storage of additional calories in the WAT fails [8], pathological ectopic deposition of fat extends to the liver [9], pancreas [9], muscle [10], and kidney [11]. Excess fat deposition in these organs renders them also resistant to insulin action and results in excessive free fatty acid release from the WAT, increased hepatic glucose release, higher fasting glucose, and postprandial hyperglycemia [12]. Non-alcoholic fatty liver disease [13] and steatohepatitis [14] result from lipotoxic fat accumulation in the liver. Possibly the most damaging concomitant of obesity in developed countries is the progression from insulin resistance and pre-diabetes to Type-2 diabetes (T2D). The incidence of T2D has increased from 4.4 million or 2.4% of the US population in 1970s [15] to 29.1

million or 9.3% of the population in 2014 [16]. Hyperglycemia and compensatory hyperinsulinemia [17] associated with insulin resistance and glucose intolerance lead to pathological glycation of circulating proteins and formation of advanced glycation end products [18]. This progression ultimately leads to a pancreatic beta cells secretory failure [19] and apoptosis [20]. Insulin-resistant and diabetic muscle has reduced capacity to store glycogen [21] and take up glucose [22]. Insulin resistance and T2D lead to endothelial dysfunction [5] and microvascular pathologies including diabetic retinopathy [23], nephropathy [24], and neuropathy [25]. At the cellular level, the progression from insulin resistance to diabetes is accompanied by oxidative stress [26] and systemic inflammation [5]. Treatment of these obesity- and diabetes-associated pathologies has engaged medical practice, burdened the afflicted individuals psychologically [27] and physiologically [28], and imposed a \$245 billion health care financial burden [29].

Medicine and associated sciences dealing with the pathologies produced by obesity and T2D have predominantly focused on the treatment rather than the prevention of the two health conditions. Both pharmacological [30-32] and surgical [33-34] approaches have been used in the treatment of both obesity and T2D. None is completely successful without incurring substantial side-effects [30,35,36], including the option of

#### Borer: Physiology for prevention of pathology

permanent changes to the gastro-intestinal anatomy and physiology. This review puts forward for consideration two ideas: (1) that some of the failures in addressing prevention of obesity and T2D are the result of incorrect characterization of the neuroendocrine control of feeding and regulation of body weight, and (2) that there is a knowledge gap in how the capacity of the pancreas and adipose tissue, key organs in the pathologies of obesity and T2D, may be modified during two important periods of tissue proliferation, the second trimester of fetal development, the pre-pubertal growth spurt, and also possibly during recovery from substantial weight loss.

#### INCORRECT CHARACTERIZATION OF THE CONTROLS OF FEEDING AND THE REGULATION OF BODY WEIGHT

At least three widely-held hypotheses about the regulation of body weight may set back the development of improved preventive or corrective strategies. They are (1) the view that regulation of body weight is homeostatic or self-correcting by way of negative feedbacks which may convey a false confidence that this mechanism will guide the body mass to a healthy set point; (2) the position that body weight regulation is based on direct negative feedback from the WAT through secretion and actions of adipokine leptin such that leptin reduces adiposity by suppressing appetite and food intake and increasing thermogenesis; and (3) the concept that the regulation of body weight depends on the integrity of leptin actions in the brain and is inoperative in its absence. These three premises are not supported by experimental data. In discussing their inadequacies, the following three sections propose an alternative physiological explanation of the sequelae of obesity and a weight-regulatory schema that is supported by experimental data.

#### Body Weight Regulation is a Consequence of Nonhomeostatic, Rather than Homeostatic, Feeding Controls and of Non-homeostatic Motivation for Physical Activity

Regulation of body weight stability entails defenses against its loss through compensatory adjustments in feeding, energy expenditure through thermogenesis and physical activity. A prerequisite for a homeostatic regulation of body weight would be the ability to sense calorie deficits generated by food restriction or energy expenditure of physical activity, and energy gain through calories eaten. There is ample evidence that within the context of daily meal-to-meal eating and episodes of physical activity, humans do not have the ability to track calories that are eaten, missing, or expended. Calories missing in the morning meal or expended during exercise are not made up during a subsequent meal [37]. The same volume of food is eaten when the opportunity for normal levels of activity is constrained [38], or when meals of different energy content are fed over an 11 weeks period [39]. The amount of food eaten is guided by food palatability [40,41], the opportunistic variables of quantity of food and drink offered [42,43], and social facilitation [44], but not the ability to track food calories [37]. The non-homeostatic nature of food intake leads to obesity as a result of exposure to "cafeteria diets" first demonstrated in

rats given a variety of highly palatable and energy-dense foods in addition to their standard chow [45]. Eating is terminated by a sensation of fullness mediated by stretch receptors in the stomach wall [46,47] and relayed to the hindbrain nucleus of the tractus solitarius. The progressive rise in hunger during the inter-meal intervals reflects diminishing absorption of nutrients in the intestines. The intestines can sense nutrient quality [47] and affect the rate of gastric emptying and nutrient transit through secretion of several gut hormones [48].

Energy expenditure of physical activity contributes to the regulated weight plateau only if it is externally mandated. The weight of humans engaged in physically-demanding occupations is lower than those in more sedentary occupation [49]. Provided with a running wheel, rats engage in spontaneous running, and their weight stabilizes at a lower level than in caged animals not having this option [50]. Spontaneous physical activity, like feeding, also is structured non-homeostatically [51]. Despite contrary expectation, the motivation for physical activity increases with weight loss. Rats provided with a running wheel, and insufficient amount of food will increase their running in parallel with weight loss to the point of inanition [52]. Likewise, anorexic humans display a "drive to be active" [53]. By contrast, non-basal energy expenditure in humans declines in proportion to increases in adiposity [54,55] with almost complete inactivity in morbid obesity [56]. That this is an issue of the interaction between adiposity, and the motivation to move can be demonstrated by providing the negative external motivation. A foot shock at the base of a treadmill equalizes the duration and intensity or forced running in overweight and lean hamsters [57].

So how do non-homeostatic controls of feeding and physical activity achieve defended stability of body weight? They do so by having a functional connection within the framework of intermittent meal eating and movement. Episodes of intermittent opportunistic food intake lead to fullness and are associated with temporary suppression of motivation to move. Completion of meal processing reactivates the motivation to move and seek and initiate another meal. Depending on the amount of physical work required to obtain the resources for food, and on the quality, quantity and palatability of available food, the weight will stabilize at different regulated weight plateaus [49].

The insight that we are unable to track calories eaten or expended and are vulnerable to overeating palatable and abundant food while living in an environment largely devoid of the need for much physical work should guide individual decisions about how much to eat and move to maintain a healthy weight plateau. The additional insight that our motivation for physical activity depends on our adiposity should serve as a helpful cue to counteract non-homeostatic tendency to overeat and to engage in healthy levels of physical activity.

#### Meal-Associated Release of Gastric Leptin is the More Likely Contributor to Acute Energy Regulation than Adiposity-Associated Leptin Release from the WAT

The current and prevailing view of the weight-regulatory mechanism is that adipokine leptin secreted from the

subcutaneous WAT acts on the arcuate (ARC) and ventromedial hypothalamic (VMH) nuclei in a negative feedback fashion to influence brain circuits that inhibit feeding and increase energy expenditure [58-63]. This hypothesis, therefore, predicts reduced food intake and adiposity as fasting leptin concentrations rise in parallel with body fat level to inhibit feeding and reduce body fat. Fasting leptin [64] as well as insulin [65] concentration do rise in parallel with the increase in body fat [Figure 1]. This prompts a misinterpretation of a cause-and-effect relationship. This hypothesis is further strengthened by the observation of increased food intake that leads to obesity in animals [66] and humans [67] unable to secrete leptin, and by correction of both pathologies by administration of leptin [66,68]. This hypothesis has achieved the status of a dogma despite the following contradictory evidence. First, administration of a range of physiological and pharmacological doses of leptin to obese humans did not suppress their food intake or reduce their adiposity [69]. Second, high-fat diets [70] and cafeteria diets [45] easily induce obesity in animals and humans [71] despite the parallel increases in fasting leptin concentration and in adiposity. Moreover, third, and most damaging to the negative-feedback formulation of the hypothesis, leptin effectiveness in suppressing food intake and adiposity is inversely rather than proportionally related to body fat [72]. Finally in the daily episodic feeding circumstances, circulating concentrations of leptin change in response to the calories eaten or calories expended in exercise but bear no relationship to sensations of hunger, fullness, or the amount of subsequent food consumed [37].

An alternative hypothesis for the role of leptin in energy regulation is based on the evidence that in the context of intermittent meal eating, leptin is secreted by the chief cells in the gastric mucosa [73,74]. Postprandial leptin secretion and actions are yoked to meal-associated insulin secretion and actions in counter-regulatory fashion [75] [Figure 2]. The well-known postprandial insulin release that is sensitive to both calories ingested by mouth and calories infused intravenously [37] upregulates within 3-4 h the production and release of leptin [37,76]. Besides its exocrine release subsequent to its forming a complex with its soluble receptor to protect it from gastric acid, gastric leptin also reaches the systemic circulation. It does so



**Figure 1:** The proportional relationship between body adiposity and fasting concentrations of fasting serum insulin (left) and fasting serum leptin. The relationship reflects progressive loss of tissue sensitivity to the two hormones that changes in parallel with the loss of receptors of the two hormones on the target cells. Data for insulin from reference 65 and for leptin from reference 64

by being transported to the duodenum where it binds with its receptor on the luminal membrane from where it is transcytozed to the Golgi apparatus of the duodenal enterocyte. There it again binds with its receptor and leaves intestinal mucosa for systemic circulation [77]. In opposition to the anabolic and parasympathetic actions of insulin to facilitate cellular uptake and storage of nutrients and to suppress mobilization and utilization of these nutrients, leptin counter regulates insulin actions by blocking its release [78] and blocking insulin binding to its receptors [79]. Leptin also increases lipolysis and lipid utilization [80,81] by mobilizing lipids stored in the adipose tissue [81], liver [82], and the muscle [83,84]. It therefore contributes to meal-to-meal balancing of positive energy balance caused by food intake, in contrast to hormones such as catecholamines, glucagon, cortisol, and growth hormone that are recruited for production of emergency fuels in response to negative energy balance. In addition to diurnal leptin secretion in response to the postprandial insulin stimulus, leptin also exhibits a circadian pattern of secretion [85] that is also sensitive to energy balance. The acrophase of circadian leptin rhythm is in the middle of the night [85] and increases after excess diurnal energy intake and declines after diurnal energy deficit [86].

The counter-regulatory relationship of leptin and postprandial insulin helps explain the paradoxical relationship of leptin to body fat as postulated by the homeostatic concept of weight regulation. The fasting concentrations of the two hormones change in lockstep with increases in body fat mass [Figure 1]. The increases in the fasting concentrations of both insulin and leptin reflect increases in tissue resistance to their actions. This is a consequence of the fundamental endocrine principle that the sensitivity of peripheral tissues to fasting concentration of a hormone declines in parallel with the number of hormone receptors on the target cell surface [Figure 3] [87] and with the degree of repletion of target tissues with storage fuels. As the amount of lipids and glycogen in the WAT [88], muscle [89,90], and liver [91] increases, the number of insulin receptors on their cell surfaces declines. At equal Km, tissue sensitivity to hormone increases in proportion to the number of "spare receptors" above the number required to elicit a biological response [Figure 3a] [87]. With a smaller number of receptors in energy-replete cells, resistance to a hormone manifests in the form of a higher hormone concentration required to elicit the biological response [Figure 3b].

The parallel rise in the fasting concentrations of insulin and leptin represents the cellular mechanism contributing to the regulation of body weight. Through increased sensitivity to insulin, triglyceride-depleted adipocytes, and lipid- and glycogen-depleted liver and skeletal muscle, are more responsive to insulin actions leading to heightened nutrient uptake and storage. Moreover, the counter-regulation of insulin by leptin controls the amount of fat storage in tissues and thus contributes to the maintenance of insulin sensitivity [80,92]. The pathologies observed in obesity and T2D simply reflect the operation of insulin and leptin above their physiological range. There is a need for systematic study of dietary and exercise conditions that reduce postprandial insulin and enhance postprandial leptin action. Such data could then inform and



**Figure 2:** The counter regulatory relationship between postprandial pancreatic insulin secretion and action and postprandial gastric leptin secretion and action. Insulin secretion associated with meal eating upregulates production and secretion of gastric leptin. Gastric leptin inhibits insulin secretion, its binding to insulin receptors, and counteracts insulin's anabolic actions

guide adjustments in feeding and activity behaviors to better match energy intake and expenditure for the maintenance of healthy body weight level.

#### Weight Regulation Depends on the Interaction of the Sympathetic and Parasympathetic Components of the Autonomic Nervous System and Not on the Integrity of Leptin Signaling

Several decades ago, the autonomic component of the central nervous system (ANS) was recognized as the principal regulatory agent for balancing energy intake and expenditure [93]. Lesions of the ARC and VMH nuclei appeared to shift the balance between the sympathetic energy expending ANS division in favor of the parasympathetic ANS division responsible for over secretion of insulin and obesity [93,94] both of which were preventable by transection of the parasympathetic nerve vagus [94]. Since then, additional evidence accumulated that ANS also controls circadian and ultradian periodicities of meal taking, food processing, and controls thermogenesis and spontaneous physical activity [75,95]. Since the discovery the capacity of leptin to suppress feeding and adiposity in animals [66] and humans [68] unable to produce leptin, the research and interpretive focus has shifted from the regulatory role of ANS to the analysis of leptin actions on the hypothalamic and midbrain circuits regulating feeding and thermogenesis [96] with the implicit hypothesis that the integrity of the leptin signaling in the brain is the essential prerequisite for the regulation of body weight. The assumption that weight regulation does not operate in the absence of functional leptinbrain interactions is not supported by facts. Animals with the lesions of the ARC-VMH hypothalamic targets of leptin action continue to defend their elevated weight and fat plateau against losses [97]. Furthermore, if animals are rendered obese prior to the damage of the VMH-ARC targets of leptin action, they



**Figure 3:** The sensitivity of peripheral tissues to fasting concentration of a hormone declines in parallel with the number of receptors on the target cell surface and the degree of repletion of target tissues with storage fuels. As the amount of the storage fuel in the target cell increases, the number of insulin receptors on the cell surfaces declines. At equal km, tissue sensitivity to a hormone increases in proportion to the number of "spare receptors" above the number required to elicit a biological response (a). With smaller number of receptors in energy replete cells, resistance to a hormone manifests in the form of a higher hormone concentration needed to elicit the biological response (b). From reference 87, with permission of the Oxford University Press

do not overeat or over secrete insulin. Instead, they maintain their elevated fat plateau without hyperphagia or insulin over secretion [98] suggesting that removal of leptin action in the brain increases the level at which the body fat is maintained but does not abolish the defense mechanism against body weight loss. So while destruction of some brain targets of leptin action increases adiposity, it does not remove the role of the ANS in regulating the higher weight and fat setpoint.

The early interpretation of actions of leptin in the brain as a direct and proportional regulator of adiposity through adjustments of feeding and thermogenesis is slowly giving way to the recognition that within their physiologic concentrations, leptin and insulin suppress the reward value or salience of food [40,41] as well as the motivation to be physically active [99,100]. A decline in their concentrations increases the strength of these motivations. Whether the brain substrates over which leptin and insulin exert these suppressive effects are confined to nucleus accumbens in the ventral striatum [40,41] or also include some hypothalamic nuclei [101] remains to be worked out. This reinterpretation of the actions of insulin and leptin in the brain dovetails with the non-homeostatic control of feeding and physical activity in weight regulation. By virtue of the changes in hormone sensitivity as a function of adiposity [Figure 1], the withdrawal of insulin and leptin in underweight state facilitates feeding and physical work to procure food as well increases the efficiency of energy storage when the metabolic fuel stores are depleted and peripheral and brain sensitivity to the two hormones is maximal. As the compensatory eating and enhanced efficiency of food storage restore the WAT lipid stores and muscle lipid and glycogen stores, rising tissue resistance to insulin and leptin actions and the suppression by higher concentrations of the two hormones of the motivational brain substrates stabilizes body weight at the pre-deprivation level. The ultimate understanding of the weight regulatory mechanism will require a shift from the current preoccupation with leptin action on hypothalamic circuits to the analysis of the role of ANS in balancing parasympathetic and sympathetic controls over secretion insulin and leptin and their actions on metabolism.

#### HOW TO INFLUENCE THE PHYSIOLOGY IN ORDER TO PREVENT THE PATHOLOGY

The obvious solutions to avoiding the pathologies that result from overeating and obesity and that lead to T2D would require implementation of social policies to reduce easy availability of highly palatable foods and to impose requirements for greater physical work. Obviously, such utopian plans will not be feasible in open-market democratic societies. That it can be achieved in the context of a totalitarian society experiencing food shortages is illustrated by the example of Cuba, where food intakes, body weights, and pathologies associated with obesity and T2D were minimized through reduced access to food and increased physical work requirements [102].

An alternative to pharmacological and surgical treatments of obesity - and T2D-associated pathologies would be to close the knowledge gap on how body fatness and pancreatic beta cell capacities could be influenced during developmental growth. The most damaging pathologies of obesity in adulthood are the result of exceeding the capacity of the adipose tissue for fat storage which leads to ectopic fat accumulation and lipotoxicity in the liver, pancreas and other tissues [103]. The most damaging pathologies of adult T2D are the consequence of hyperglycemia and compensatory hyper-insulinemia leading to progressive apoptosis of pancreatic beta cells. There is a lack of knowledge whether modifications of nutrition and exercise could epigenetically modify the proliferative capacity of pancreatic beta cells and WAT adipocytes to resist the damage inflicted by overeating and excessive storage of nutrient energy. The likely windows of opportunity for epigenetic modification of these two tissues include the periods of rapid cellular proliferation during the second trimester of intrauterine growth and during the pre-pubertal growth spurt [104]. It is not clear why the negative energy balance during pregnancy generated by dietary restriction and exercise produces opposite effects. Dietary restriction during later stages of pregnancy reduces overall fetal growth and organ size [105-109]. Offspring subjected to intrauterine growth retardation have reduced pancreatic  $\beta$  cell mass [105-108], increased fasting insulin, reduced glucose tolerance [108], and increased hepatic insulin resistance [109]. On the other hand, exercise energy expenditure during pregnancy which also reduces overall offspring growth and body fat measured at birth [110,111], at 1-year of age [110] and at 5 years of age [111] affects glucose tolerance and insulin sensitivity in the opposite way. Infants of exercising mothers weight about 6-7% less than of non-exercising pregnant women, are about 7% shorter, and have 25-31% less body fat [110,111]. At age 5, their subcutaneous skinfold measurements are 16% lower than those in offspring of sedentary mothers [111]. If forced to swim, weanling rats (which are born in less mature state than human infants), display reduced adipocyte cellularity [112]. Yet despite reduced body and individual organ size at birth as a result of maternal exercise during pregnancy, such rodents in adulthood display improved glucose tolerance [113-115], and systemic insulin sensitivity measured by hyperinsulinemic hyperglycemic clamp [114]. Why energy restriction during pregnancy by dietary and exercise means produce opposite results is not known. Nor is it known why children of obese mothers, as well as children of mothers experiencing dietary restriction during pregnancy, experience increased risk of insulin resistance and T2D [116]. Clearly, this is an area in need of systematic research to increase the understanding how exercise and diets may affect pancreatic and WAT development.

Another window of opportunity for potential epigenetic changes in body composition and pancreatic beta cell capacity may be the period during maintenance of, and recovery from, dietinduced weight loss. Rats exposed to exercise during weight-loss maintenance display reduced rate of weight regain during ad libitum re-alimentation [117]. Leptin treatment during weightloss maintenance in humans reduces hunger and increases thermogenesis [118]. Leptin treatment during weight loss maintenance reduced body fat regain in non-growing hamsters and repartitioned body composition in favor of lean body mass in growing hamsters [119]. That leptin could produce epigenetic changes in the brain targets of its action is seen in functional enhancement of the activity of brain areas involved in detecting the salience and rewarding value of food during fasting in three hypoleptinemic subjects treated with leptin [120]. The knowledge gap regarding whether manipulations reducing WAT cellularity are beneficial or increase insulin resistance as is the case in lipodystrophy also needs to be closed.

The ideas proposed in this review are intended to encourage a research focus toward (1) a better understanding of the mechanism of human feeding and locomotion to empower individuals with information on how to prevent overeating and excessive weight gain; (2) an understanding of the counter regulatory relationship between insulin and leptin in the context of meal-to-meal eating to produce data that could define optimal relationship between calories eaten and expended; and (3) closing the information gap regarding the influence of quantities and qualities of nutrients and quantity and type of exercise during growth periods and recovery from weight loss when the cellularity of adipose and pancreatic endocrine tissues may be sensitive to epigenetic modification. This type of research would augment the understanding of physiology and reduce the burden of the pathological consequences of obesity and T2D.

#### REFERENCES

- Available from: http://www.cdc.gov/obesity/data/adult.htm. [Last accessed on 2015 May 18].
- Vanecková I, Maletínská L, Behuliak M, Nagelová V, Zicha J, Kuneš J. Obesity-related hypertension: Possible pathophysiological

mechanisms. J Endocrinol 2014;223:R63-78.

- Lovren F, Teoh H, Verma S. Obesity and atherosclerosis: Mechanistic insights. Can J Cardiol 2015;31:177-83.
- Samad F, Ruf W. Inflammation, obesity, and thrombosis. Blood 2013;122:3415-22.
- Iantorno M, Campia U, Di Daniele N, Nistico S, Forleo GB, Cardillo C, et al. Obesity, inflammation and endothelial dysfunction. J Biol Regul Homeost Agents 2014;28:169-76.
- Rankinen T, Sarzynski MA, Ghosh S, Bouchard C. Are there genetic paths common to obesity, cardiovascular disease outcomes, and cardiovascular risk factors? Circ Res 2015;116:909-22.
- 7. Samuel VT, Petersen KF, Shulman GI. Lipid-induced insulin resistance: Unravelling the mechanism. Lancet 2010;375:2267-77.
- Shulman GI. Ectopic fat in insulin resistance, dyslipidemia, and cardiometabolic disease. N Engl J Med 2014;371:1131-41.
- van der Zijl NJ, Goossens GH, Moors CC, van Raalte DH, Muskiet MH, Pouwels PJ, *et al.* Ectopic fat storage in the pancreas, liver, and abdominal fat depots: Impact on ß-cell function in individuals with impaired glucose metabolism. J Clin Endocrinol Metab 2011;96:459-67.
- Taira S, Shimabukuro M, Higa M, Yabiku K, Kozuka C, Ueda R, *et al.* Lipid deposition in various sites of the skeletal muscles and liver exhibits a positive correlation with visceral fat accumulation in middle-aged Japanese men with metabolic syndrome. Intern Med 2013;52:1561-71.
- Guebre-Egziabher F, Alix PM, Koppe L, Pelletier CC, Kalbacher E, Fouque D, *et al.* Ectopic lipid accumulation: A potential cause for metabolic disturbances and a contributor to the alteration of kidney function. Biochimie 2013;95:1971-9.
- 12. Taylor R. Banting Memorial lecture 2012: Reversing the twin cycles of type 2 diabetes. Diabet Med 2013;30:267-75.
- Than NN, Newsome PN. A concise review of non-alcoholic fatty liver disease. Atherosclerosis 2015;239:192-202.
- Neuman MG, French SW, French BA, Seitz HK, Cohen LB, Mueller S, et al. Alcoholic and non-alcoholic steatohepatitis. Exp Mol Pathol 2014;97:492-510.
- Fox CS, Pencina MJ, Meigs JB, Vasan RS, Levitzky YS, D'Agostino RB Sr. Trends in the incidence of type 2 diabetes mellitus from the 1970s to the 1990s: The framingham heart study. Circulation 2006;113:2914-8.
- National Diabetes Statistics Report, 2014. Available from: http://www. cdc.gov/diabetes/pubs/statsreport14/national-diabetes-report-web. pdf. [Last accessed on 2015 May 18].
- DeFronzo RA. Lilly lecture 1987. The triumvirate: Beta-cell, muscle, liver. A collusion responsible for NIDDM. Diabetes 1988;37:667-87.
- Nowotny K, Jung T, Höhn A, Weber D, Grune T. Advanced glycation end products and oxidative stress in type 2 diabetes mellitus. Biomolecules 2015;5:194-222.
- Halban PA, Polonsky KS, Bowden DW, Hawkins MA, Ling C, Mather KJ, *et al.* ß-cell failure in type 2 diabetes: Postulated mechanisms and prospects for prevention and treatment. Diabetes Care 2014;37:1751-8.
- 20. Anuradha R, Saraswati M, Kumar KG, Rani SH. Apoptosis of beta cells in diabetes mellitus. DNA Cell Biol 2014;33:743-8.
- 21. Beck-Nielsen H. The role of glycogen synthase in the development of hyperglycemia in type 2 diabetes: 'To store or not to store glucose, that's the question'. Diabetes Metab Res Rev 2012;28:635-44.
- Abdul-Ghani MA, DeFronzo RA. Pathogenesis of insulin resistance in skeletal muscle. J Biomed Biotechnol 2010;2010:476279.
- Aiello LM. Perspectives on diabetic retinopathy. Am J Ophthalmol 2003; 136:122-35.
- Maezawa Y, Takemoto M, Yokote K. Cell biology of diabetic nephropathy: Roles of endothelial cells, tubulointerstitial cells and podocytes. J Diabetes Investig 2015;6:3-15.
- Callaghan BC, Cheng HT, Stables CL, Smith AL, Feldman EL. Diabetic neuropathy: Clinical manifestations and current treatments. Lancet Neurol 2012;11:521-34.
- 26. Paneni F, Costantino S, Cosentino F. Role of oxidative stress in endothelial insulin resistance. World J Diabetes 2015;6:326-32.
- Chew BH, Shariff-Ghazali S, Fernandez A. Psychological aspects of diabetes care: Effecting behavioral change in patients. World J Diabetes 2014;5:796-808.
- Seaquist ER. Addressing the burden of diabetes. JAMA 2014;311:2267-8.

- 29. American Diabetes Association. Economic costs of diabetes in the U.S. in 2012. Diabetes Care 2013;36:1033-46.
- Kakkar AK, Dahiya N. Drug treatment of obesity: Current status and future prospects. Eur J Intern Med 2015;26:89-94.
- Hurt RT, Edakkanambeth Varayil J, Ebbert JO. New pharmacological treatments for the management of obesity. Curr Gastroenterol Rep 2014;16:394.
- 32. Kahn SE, Buse JB. Medications for type 2 diabetes: How will we be treating patients in 50 years? Diabetologia 2015.
- Boido A, Ceriani V, Cetta F, Lombardi F, Pontiroli AE. Bariatric surgery and prevention of cardiovascular events and mortality in morbid obesity: Mechanisms of action and choice of surgery. Nutr Metab Cardiovasc Dis 2015;25:437-43.
- 34. Tham JC, Howes N, le Roux CW. The role of bariatric surgery in the treatment of diabetes. Ther Adv Chronic Dis 2014;5:149-57.
- Azimova K, San Juan Z, Mukherjee D. Cardiovascular safety profile of currently available diabetic drugs. Ochsner J 2014;14:616-32.
- Tack J, Deloose E. Complications of bariatric surgery: Dumping syndrome, reflux and vitamin deficiencies. Best Pract Res Clin Gastroenterol 2014;28:741-9.
- Borer KT, Wuorinen E, Ku K, Burant C. Appetite responds to changes in meal content, whereas ghrelin, leptin, and insulin track changes in energy availability. J Clin Endocrinol Metab 2009;94:2290-8.
- Shepard TY, Weil KM, Sharp TA, Grunwald GK, Bell ML, Hill JO, *et al.* Occasional physical inactivity combined with a high-fat diet may be important in the development and maintenance of obesity in human subjects. Am J Clin Nutr 2001;73:703-8.
- Kendall A, Levitsky DA, Strupp BJ, Lissner L. Weight loss on a low-fat diet: Consequence of the imprecision of the control of food intake in humans. Am J Clin Nutr 1991;53:1124-9.
- Berridge KC. Food reward: Brain substrates of wanting and liking. Neurosci Biobehav Rev 1996;20:1-25.
- Peciña S, Berridge KC. Hedonic hot spot in nucleus accumbens shell: Where do mu-opioids cause increased hedonic impact of sweetness? J Neurosci 2005;25:11777-86.
- Smarandescu L, Walker D, Wansink B. Mindless drinking: How gender and BMI relate to the consumption of alcohol. Int J Drug Policy 2014;25:1131-4.
- van Kleef E, Shimizu M, Wansink B. Serving bowl selection biases the amount of food served. J Nutr Educ Behav 2012;44:66-70.
- De Castro JM. Social facilitation of duration and size but not rate of the spontaneous meal intake of humans. Physiol Behav 1990;47:1129-35.
- Sclafani A, Springer D. Dietary obesity in adult rats: Similarities to hypothalamic and human obesity syndromes. Physiol Behav 1976;17:461-71.
- 46. Phillips RJ, Powley TL. Gastric volume rather than nutrient content inhibits food intake. Am J Physiol 1996;271:R766-9.
- 47. Powley TL, Phillips RJ. Gastric satiation is volumetric, intestinal satiation is nutritive. Physiol Behav 2004;82:69-74.
- Seimon RV, Lange K, Little TJ, Brennan IM, Pilichiewicz AN, Feltrin KL, et al. Pooled-data analysis identifies pyloric pressures and plasma cholecystokinin concentrations as major determinants of acute energy intake in healthy, lean men. Am J Clin Nutr 2010;92:61-8.
- Mayer J, Roy P, Mitra KP. Relation between caloric intake, body weight, and physical work: Studies in an industrial male population in West Bengal. Am J Clin Nutr 1956;4:169-75.
- Tokuyama K, Saito M, Okuda H. Effects of wheel running on food intake and weight gain of male and female rats. Physiol Behav 1982;28:899-903.
- Borer KT. Nonhomeostatic control of human appetite and physical activity in regulation of energy balance. Exerc Sport Sci Rev 2010;38:114-21.
- Routtenberg A, Kuznesof AW. Self-starvation of rats living in activity wheels on a restricted feeding schedule. J Comp Physiol Psychol 1967;64:414-21.
- 53. Casper RC. The 'drive for activity' and "restlessness" in anorexia nervosa: Potential pathways. J Affect Disord 2006;92:99-107.
- Rising R, Harper IT, Fontvielle AM, Ferraro RT, Spraul M, Ravussin E. Determinants of total daily energy expenditure: Variability in physical activity. Am J Clin Nutr 1994;59:800-4.
- Schulz LO, Schoeller DA. A compilation of total daily energy expenditures and body weights in healthy adults. Am J Clin Nutr 1994;60:676-81.
- 56. Vanhecke TE, Franklin BA, Miller WM, deJong AT, Coleman CJ,

McCullough PA. Cardiorespiratory fitness and sedentary lifestyle in the morbidly obese. Clin Cardiol 2009;32:121-4.

- Borer KT, Potter CD, Fileccia N. Basis for the hypoactivity that accompanies rapid weight gain in hamsters. Physiol Behav 1983;30:389-97.
- Schwartz MW, Woods SC, Porte D Jr, Seeley RJ, Baskin DG. Central nervous system control of food intake. Nature 2000;404:661-71.
- Woods SC, Schwartz MW, Baskin DG, Seeley RJ. Food intake and the regulation of body weight. Annu Rev Psychol 2000;51:255-77.
- Benoit SC, Clegg DJ, Seeley RJ, Woods SC. Insulin and leptin as adiposity signals. Recent Prog Horm Res 2004;59:267-85.
- 61. Woods SC, D'Alessio DA. Central control of body weight and appetite. J Clin Endocrinol Metab 2008;93:S37-50.
- Ryan KK, Woods SC, Seeley RJ. Central nervous system mechanisms linking the consumption of palatable high-fat diets to the defense of greater adiposity. Cell Metab 2012;15:137-49.
- Guyenet SJ, Schwartz MW. Clinical review: Regulation of food intake, energy balance, and body fat mass: Implications for the pathogenesis and treatment of obesity. J Clin Endocrinol Metab 2012;97:745-55.
- Considine RV, Sinha MK, Heiman ML, Kriauciunas A, Stephens TW, Nyce MR, *et al.* Serum immunoreactive-leptin concentrations in normal-weight and obese humans. N Engl J Med 1996;334:292-5.
- 65. Bagdade JD. Bagdade JD: Basal insulin and obesity. Lancet 1968;2:630-1.
- Pelleymounter MA, Cullen MJ, Baker MB, Hecht R, Winters D, Boone T, *et al.* Effects of the obese gene product on body weight regulation in ob/ob mice. Science 1995;269:540-3.
- Montague CT, Farooqi IS, Whitehead JP, Soos MA, Rau H, Wareham NJ, *et al.* Congenital leptin deficiency is associated with severe early-onset obesity in humans. Nature 1997;387:903-8.
- Farooqi IS, Jebb SA, Langmack G, Lawrence E, Cheetham CH, Prentice AM, *et al.* Effects of recombinant leptin therapy in a child with congenital leptin deficiency. N Engl J Med 1999;341:879-84.
- Heymsfield SB, Greenberg AS, Fujioka K, Dixon RM, Kushner R, Hunt T, *et al.* Recombinant leptin for weight loss in obese and lean adults: A randomized, controlled, dose-escalation trial. JAMA 1999;282:1568-75.
- Lemonnier D, Suquet JP, Aubert R, De Gasquet P, Pequignot E. Metabolism of the mouse made obese by a high-fat diet. Diabete Metab 1975;1:77-85.
- Hovell MF, Mewborn CR, Randle Y, Fowler-Johnson S. Risk of excess weight gain in university women: A three-year community controlled analysis. Addict Behav 1985;10:15-28.
- Frederich RC, Hamann A, Anderson S, Löllmann B, Lowell BB, Flier JS. Leptin levels reflect body lipid content in mice: Evidence for diet-induced resistance to leptin action. Nat Med 1995;1:1311-4.
- Sobhani I, Bado A, Vissuzaine C, Buyse M, Kermorgant S, Laigneau JP, et al. Leptin secretion and leptin receptor in the human stomach. Gut 2000;47:178-83.
- Cammisotto PG, Renaud C, Gingras D, Delvin E, Levy E, Bendayan M. Endocrine and exocrine secretion of leptin by the gastric mucosa. J Histochem Cytochem 2005;53:851-60.
- Borer KT. Counterregulation of insulin by leptin as key component of autonomic regulation of body weight. World J Diabetes 2014;5:606-29.
- Utriainen T, Malmström R, Mäkimattila S, Yki-Järvinen H. Supraphysiological hyperinsulinemia increases plasma leptin concentrations after 4 h in normal subjects. Diabetes 1996;45:1364-6.
- Cammisotto PG, Levy E, Bukowiecki LJ, Bendayan M. Cross-talk between adipose and gastric leptins for the control of food intake and energy metabolism. Prog Histochem Cytochem 2010;45:143-200.
- Ishida K, Murakami T, Mizuno A, Iida M, Kuwajima M, Shima K. Leptin suppresses basal insulin secretion from rat pancreatic islets. Regul Pept 1997;70:179-82.
- Walder K, Filippis A, Clark S, Zimmet P, Collier GR. Leptin inhibits insulin binding in isolated rat adipocytes. J Endocrinol 1997;155:R5-7.
- Barzilai N, Wang J, Massilon D, Vuguin P, Hawkins M, Rossetti L. Leptin selectively decreases visceral adiposity and enhances insulin action. J Clin Invest 1997;100:3105-10.
- 81. Harris RB. Direct and indirect effects of leptin on adipocyte metabolism. Biochim Biophys Acta 2014;1842:414-23.
- Huang W, Dedousis N, O'Doherty RM. Hepatic steatosis and plasma dyslipidemia induced by a high-sucrose diet are corrected by an acute leptin infusion. J Appl Physiol (1985) 2007;102:2260-5.

- Todd MK, Yaspelkis BB 3<sup>rd</sup>, Turcotte LP. Short-term leptin treatment increases fatty acids uptake and oxidation in muscle of high fat-fed rats. Metabolism 2005;54:1218-24.
- Steinberg GR, Parolin ML, Heigenhauser GJ, Dyck DJ. Leptin increases FA oxidation in lean but not obese human skeletal muscle: Evidence of peripheral leptin resistance. Am J Physiol Endocrinol Metab 2002;283:E187-92.
- Wardlaw SL, Burant CF, Klein S, Meece K, White A, Kasten T, *et al.* Continuous 24-hour leptin, proopiomelanocortin, and amino acid measurements in human cerebrospinal fluid: Correlations with plasma leptin, soluble leptin receptor, and amino acid levels. J Clin Endocrinol Metab 2014;99:2540-8.
- van Aggel-Leijssen DP, van Baak MA, Tenenbaum R, Campfield LA, Saris WH. Regulation of average 24h human plasma leptin level; The influence of exercise and physiological changes in energy balance. Int J Obes Relat Metab Disord 1999;23:151-8.
- Mendelson CR. Mechanisms of hormone action. In: Textbook of Endocrine Physiology. 3<sup>rd</sup> ed. New York: Oxford University Press; 1996. p. 32.
- Olefsky JM, Kobayashi M. Mechanisms of fasting-induced increase in insulin binding to rat adipocytes. J Clin Invest 1978;61:329-38.
- Brady LJ, Goodman MN, Kalish FN, Ruderman NB. Insulin binding and sensitivity in rat skeletal muscle: Effect of starvation. Am J Physiol 1981;240:E184-90.
- Le Marchand-Brustel Y, Jeanrenaud B, Freychet P. Insulin binding and effects in isolated soleus muscle of lean and obese mice. Am J Physiol 1978;234:E348-58.
- Fleig WE, Enderle D, Steudter S, Nöther-Fleig G, Ditschuneit H. Regulation of basal and insulin-stimulated glycogen synthesis in cultured hepatocytes. Inverse relationship to glycogen content. J Biol Chem 1987;262:1155-60.
- Shimabukuro M, Koyama K, Chen G, Wang MY, Trieu F, Lee Y, *et al.* Direct antidiabetic effect of leptin through triglyceride depletion of tissues. Proc Natl Acad Sci U S A 1997;94:4637-41.
- Bray GA, York DA. Hypothalamic and genetic obesity in experimental animals: An autonomic and endocrine hypothesis. Physiol Rev 1979;59:719-809.
- Berthoud HR, Jeanrenaud B. Acute hyperinsulinemia and its reversal by vagotomy after lesions of the ventromedial hypothalamus in anesthetized rats. Endocrinology 1979;105:146-51.
- Wuorinen EC, Borer KT. Circadian and ultradian components of hunger in human non-homeostatic meal-to-meal eating. Physiol Behav 2013;122:8-16.
- 96. Gautron L, Elmquist JK. Sixteen years and counting: An update on leptin in energy balance. J Clin Invest 2011;121:2087-93.
- Brooks CM, Lambert HF. A study of the effect of limitation of food intake and the method of feeding on the rate of weight gain during hypothalamic obesity in the albino rat. Am J Physiol 1946;147:695-707.
- Hoebel BG, Teitelbaum P. Weight regulation in normal and hypothalamic hyperphagic rats. J Comp Physiol Psychol 1966;61:189-93.
- Figlewicz DP, Benoit SC. Insulin, leptin, and food reward: Update 2008. Am J Physiol Regul Integr Comp Physiol 2009;296:R9-R19.
- Figlewicz DP, Sipols AJ. Energy regulatory signals and food reward. Pharmacol Biochem Behav 2010;97:15-24.
- Myers MG Jr, Münzberg H, Leinninger GM, Leshan RL. The geometry of leptin action in the brain: More complicated than a simple ARC. Cell Metab 2009;9:117-23.
- Rodríguez-Ojea A, Jiménez S, Berdasco A, Esquivel M. The nutrition transition in Cuba in the nineties: An overview. Public Health Nutr 2002;5:129-33.
- 103. Heilbronn L, Smith SR, Ravussin E. Failure of fat cell proliferation, mitochondrial function and fat oxidation results in ectopic fat storage, insulin resistance and type II diabetes mellitus. Int J Obes Relat Metab Disord 2004;28:S12-21.
- Borer KT. Why do exercise and dietary restriction during pregnancy affect glucose tolerance in opposite ways? Diabetes 2015;64:335-7.
- Styrud J, Eriksson UJ, Grill V, Swenne I. Experimental intrauterine growth retardation in the rat causes a reduction of pancreatic B-cell mass, which persists into adulthood. Biol Neonate 2005;88:122-8.
- 106. Bertin E, Gangnerau MN, Bellon G, Bailbé D, Arbelot De Vacqueur A, Portha B. Development of beta-cell mass in fetuses of rats deprived of protein and/or energy in last trimester of pregnancy. Am J Physiol Regul Integr Comp Physiol 2002;283:R623-30.

- 107. Blondeau B, Garofano A, Czernichow P, Bréant B. Age-dependent inability of the endocrine pancreas to adapt to pregnancy: A longterm consequence of perinatal malnutrition in the rat. Endocrinology 1999;140:4208-13.
- Garofano A, Czernichow P, Bréant B. Beta-cell mass and proliferation followinglate fetal and early postnatal malnutrition in the rat. Diabetologia 1998;41:1114-20.
- Holemans K, Verhaeghe J, Dequeker J, Van Assche FA. Insulin sensitivity in adult female rats subjected to malnutrition during the perinatal period. J Soc Gynecol Investig 1996;3:71-7.
- 110. Clapp JF 3<sup>rd</sup>, Simonian S, Lopez B, Appleby-Wineberg S, Harcar-Sevcik R. The one-year morphometric and neurodevelopmental outcome of the offspring of women who continued to exercise regularly throughout pregnancy. Am J Obstet Gynecol 1998;178:594-9.
- 111. Clapp JF 3<sup>rd</sup>. Morphometric and neurodevelopmental outcome at age five years of the offspring of women who continued to exercise regularly throughout pregnancy. J Pediatr 1996;129:856-63.
- 112. Oscai LB, Babirak SP, McGarr JA, Spirakis CN. Effect of exercise on adipose tissue cellularity. Fed Proc 1974;33:1956-8.
- Carter LG, Lewis KN, Wilkerson DC, Tobia CM, Ngo Tenlep SY, Shridas P, et al. Perinatal exercise improves glucose homeostasis in adult offspring. Am J Physiol Endocrinol Metab 2012;303:E1061-8.
- Carter LG, Qi NR, De Cabo R, Pearson KJ. Maternal exercise improves insulin sensitivity in mature rat offspring. Med Sci Sports Exerc 2013;45:832-40.
- 115. Stanford KI, Lee MY, Getchell KM, So K, Hirshman MF, Goodyear LJ. Exercise before and during pregnancy prevents the deleterious effects of maternal high-fat feeding on metabolic health of male offspring. Diabetes 2015;64:427-33.

- 116. Eriksson JG, Sandboge S, Salonen MK, Kajantie E, Osmond C. Longterm consequences of maternal overweight in pregnancy on offspring later health: Findings from the Helsinki Birth Cohort Study. Ann Med 2014;46:434-8.
- 117. MacLean PS, Higgins JA, Wyatt HR, Melanson EL, Johnson GC, Jackman MR, *et al.* Regular exercise attenuates the metabolic drive to regain weight after long-term weight loss. Am J Physiol Regul Integr Comp Physiol 2009;297:R793-802.
- Rosenbaum M, Goldsmith R, Bloomfield D, Magnano A, Weimer L, Heymsfield S, *et al.* Low-dose leptin reverses skeletal muscle, autonomic, and neuroendocrine adaptations to maintenance of reduced weight. J Clin Invest 2005;115:3579-86.
- Borer KT, Devlin M, Jepsen KJ, Li L, Yang Y. Leptin re-partitions body composition during growth and recovery from weight loss. 2015; Abstract, 97<sup>th</sup> Meeting of the Endocrine Society.
- 120. Farr OM, Fiorenza C, Papageorgiou P, Brinkoetter M, Ziemke F, Koo BB, et al. Leptin therapy alters appetite and neural responses to food stimuli in brain areas of leptin-sensitive subjects without altering brain structure. J Clin Endocrinol Metab 2014;99:E2529-38.

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